AMENDMENTS TO THE SPECIFICATION

Please replace the paragraph starting on page 6, line 25 and ending on page 7, line 4 with the following amended paragraph:

Figure 3 shows alignment of the 3' UTR sequences of GBV-B. The sequence of the infectious clone of GBV-B (pGBB) is shown at the top (nts. 9038-9399 of SEQ ID NO: 1). The other sequences shown are: pGBB5-1, a non-infectious clone of GBV-B; GBV-B, a prototype of GBV-B (Simons 1995); eleven "gb" clones obtained from CT 11/91 liver homogenate by 5' RACE on the minus-strand GBV-B RNA; four "29" clones obtained from GB 2/94 pool by RT-PCR across 5'-to-3'-end-ligated viral GBV-B RNA; and seven "GBB3" clones obtained from GB 2/94 pool by standard RT-PCR. With pGBB as the reference, nucleotide substitutions or insertions are shown as uppercase letters, identical nucleotides are shown as dots and nucleotide deletions are shown as dashes.

Please replace the paragraph on page 7, lines 5-7 with the following amended paragraph:

Figure 4 shows the predicted secondary structure of the 3' UTRs of GBV-B (nts. 9038-9399 of SEQ ID NO: 1) and HCV (nts. 9363-9599 of SEQ ID NO: 6) as determined by the program "mfold" (Genetics Computer Group).

Please replace the paragraph on page 7, lines 21-24 with the following amended paragraph:

Figures 6A-6F show the nucleotide sequence (SEQ ID NO: 6) of the infectious hepatitis C virus clone of genotype 1a strain H77C and Figures 6G-6H show the amino acid sequence (SEQ ID NO: 7) encoded by the clone.

Please replace the paragraph on page 7, lines 25-28 with the following amended paragraph:

Figures 7A-7F show the nucleotide sequence (SEQ ID NO: 8) of the infectious hepatitis C virus clone of genotype 1b strain HC-J4 and Figures 7G-H show the amino acid sequence SEQ ID NO: 9) encoded by the clone.

Please replace the paragraph on page 20 line 1 through page 21 line 5 with the following amended paragraph:

Serum samples were collected weekly from the tamarins and monitored for liver enzyme levels [alanine aminotransferase (ALT), gamma-glutamyltranspeptidase (GGT), and isocitrate dehydrogenase (ICD)] by standard methods and for GBV-B RNA by a specific reverse transcriptase-polymerase chain reaction (RT-PCR) assay. Total RNA was extracted from 100 µl of serum using the TRIzol reagent. The RNA pellet was resuspended in 10 mM dithiothreitol (DTT) containing 5% (vol/vol) of RNasin (20-40 u/µl) (Promega). The RT-nested PCR was performed with primers from the 5' UTR of GBV-B (external primer pair: 5'-CCT AGC AGG GCG TGG GGG ATT TCC-3' (SEQ ID NO: 10) and 5'-AGG TCT GCG TCC TTG GTA GTG ACC-3' (SEQ ID NO: 11); internal primer pair: 5'-GGA TTT CCC CTG CCC GTC TG-3' (SEQ ID NO: 12) and 5'-CCC CGG TCT TCC CTA CAG TG-3' (SEQ ID NO: 13)). The reverse transcription was performed with avian myeloblastosis virus reverse transcriptase (Promega) and the external anti-sense primer and nested PCR was performed with AmpliTaq DNA polymerase or AmpliTaq Gold DNA polymerase (Perkin Elmer) as described previously (Bukh 1998a). Specificity was confirmed by sequence analysis of selected DNA products. Each set of experiments included a positive control sample (a 10^{-6} dilution of GB 8/93, estimated titer 100 genome equivalent (GE)) and appropriate negative control samples. The genome equivalent (GE) titer of GBV-B in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh 1998a). One GE was defined as the number of GBV-B genomes present in the highest dilution positive in RT-nested PCR. The sensitivity of this RT-nested PCR assay for GBV-B was equivalent to that of our RT-nested PCR assay for HCV (Bukh 1998b), for example, conserved NS3 primers which had the same sensitivity for GBV-B as the 5' UTR primers could detect HCV at optimal sensitivity in samples with known HCV genome titer. Testing for GBV-A and GBV-A variants was performed by RT-nested PCR assays as described previously (Bukh 1997a).

Please delete Table 1 on page 24 and substitute the following Table 1:

Table 1

Nucleotide and amino acid differences among GBV-B (Simons 1995a), the consensus sequence of GBV-B recovered from a virus pool used as the cloning source (GBV-B, 2/94) and the infectious clone of GBV-B (pGBB).

Genomic Region*	Position nt [aa]	Nucleotide GBV-B			Amino Acid GBV-B		
		GBV-B	2/94	pGBB	GBV-B	2/94	PGBB
5' UTR (1-445)							
C (446-913)							
E1 (914- 1489)	1030	C	T	T			
E2 (1490-2641)	1498	T	C (t)	C			
	1628 [395]	G	A (g)	A	V	I (V)	I
	2552 [703]	G	A (g)	A	D	N (D)	N
	2562,2563 [706]	C,A	A,C	A,C	P	H	H
	2566	T	C	C			
	2625 [727]	C	T	T	A	V	V
NS2 (2642-3385)	2647	C	T (c)	T			
	2816 [791]	C	T	T	L	F	F
	2855 [804]	A	G	G	T	A	A
	3235	A	G	G			
NS3 (3386-5125)	3475**	C	C (t)	T			
	3760	C	T (c)	T			
	4114	C	T	T			
	4117	C	A	A			
	4177	T	C	c			
	4615	C	T	T			
NS4A (5126-5290)							
NS4B (5291-6034)	5329	C	T	T			
	5332	T	C	C			
	5350	A	C	C			
	5455	C	T (c)	T			
NS5A (6035-7267)	6413 [1990]	T	A (t)	A	L	M (L)	M
	6577	G	T	T			
	6690 [2082]	T	C (t)	C	I	T(I)	T
	6965 [2174]	T	C (t)	C	S	P (S)	P
	7015	A	G (a)	G			
	7128 [2228]	G	A	A	G	E	E
	7138	A	A	G			
	7142 [2233]	A	G	G	T	A	A
NS5B (7268-9037)	7282	T	C (t)	c			
	7849	C	A	A			
	7852	C	T	T			
	8942 [2981]	G	A (g)	A	V	I (V)	I
	8971	T	C	C			

Genomic Region*	Position nt [aa]	Nucleotide GBV-B		Amino Acid GBV-B
	9026	C	T (c)	T
3' UTR (9038-9399)	9061	T	C	C
	Poly(U)	27 nts	11-23 nts	23 nts
	9134	Deletion	C	C
	9141-9399	ND	259 nts	259 nts

*Nucleotide positions corresponding to pGBB. Putative gene borders defined as suggested by homology with HCV (Muerhoff 1995). No homology was observed at the NS2- NS3 junction.
**Positions that differ between the cloning source (GBV-B 2/94) and the infectious clone of GBV-B (pGBB). The change introduced into pGBB at position 7138 introduced an artificial SalI site. nd: Not determined. Nucleotides and amino acids shown in parenthesis were found as a minor species in the cloning source (GBV-B, 2/94)